

Ecological factors influencing primate vision conformation: a phylogenetic regression workflow for the *mmodely* R-package (Version 0.2.5)

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November 7, 2024

1 Introduction

Traits evolve in the context of historically and ecologically complex arrangements that can present difficulty for researchers attempting to uncover causal relationships [5] or draw inference to larger populations [7]. Primates represent a manageably diverse clade of mammals who exhibit a wide range of behaviors and morphology conducive to revealing evolutionary processes. Ideas on the origins of primates range from predation detection [1] or deterrence [17] to targeting in hunting [2] or locomotion [3]. Here, using the *mmodely* package on primate data for locomotion [16] and vision [4], I demonstrate how the origins of primate cranial morphology can be elucidated via several ecological variables from numerous datasets [14]. Model averaging [MA] [8] and model selection [MS] [9] results primarily highlight arboreal locomotor targeting and trophic security [17] variables (such as stature or group size) as playing key roles in determining convergence of primate orbits. The *mmodely* package enables implementation of a combination of phylogenetic controlled regression [10] and information theoretic [11] (MA and MS) examination to simultaneously compare (weighted) predictor coefficients across the numerous sub-datasets generated during the exploration of all possible model combinations.

2 Licensing

The *mmodely* package is licensed under the Apache License v2.0: it is therefore free to use and redistribute, however, we, the copyright holders, wish to maintain primary control over any further development. Please be sure to cite *mmodely* if you use the package in presentations or work leading to publication.

3 Installation

This package largely depends upon the *caper* package, but most functions do not require any particular library. It is recommended that you have *caper*, *ape*, and the *caroline* package installed as a minimum.

```
> # wget https://cran.r-project.org/src/contrib/Archive/caroline/caroline_0.8.0.tar.gz
> # wget https://cran.r-project.org/src/contrib/Archive/caper/caper_0.5.tar.gz
> # wget https://cran.r-project.org/src/contrib/Archive/ape/ape_3.0-5.tar.gz
> # R CMD INSTALL caroline_0.8.0.tar.gz
> # R CMD INSTALL caper_0.5.tar.gz
> # R CMD INSTALL ape_3.0-5.tar.gz
```

Building the *mmodely* package from source requires that you have the proper dependency packages, *caroline*, installed from CRAN. This can typically be accomplished via the following commands from within the R command line environment:

```
install.packages(c('caroline','ape','caper'))
```

After a successful installation the *mmodely* package can be loaded in the normal way: by starting R and invoking the following `library` command:

```
> library(caper)
> library(mmodely)
```

4 Reading in Data

Read in the tree [15] and datasets then merge them together.

```
> data.path <- system.file("extdata","primate-example.data.csv", package="mmodely")
> data <- read.csv(data.path, row.names=1)
> data$gn_sp <- rownames(data)
> # merge data sets here if applicable
>
> tree.path <- system.file("extdata","primate-springer.2012.tre", package="mmodely")
> phyl <- ape::read.tree(tree.path)[[5]]
> #5. RAXML phylogram based on the 61199 bp concatenation of 69 nuclear and ten mitochondrial genes.
>
> phyl <- trim.phylo(phylo=phyl, gs.vect=data$gn_sp) # prune unused nodes and branches
> comp <- comp.data(phylo=phyl, df=data)
```

Typically there will be some missing data (species) in certain sources that do not occur in others. A merge of these will result in NA values for some cells. The more missing cells and merges there are, the more sub-datasets will be possible, due to case-wise deletion in the process of combinatorics underlying model iteration, averaging, and selection. The above example has little if any missing data, but the examples below introduce some artificially.

5 Basic Reporting

First, for illustration purposes, we perform a simple analysis of a single model using 'pgls' directly from the *caper* package, then show-off the 'pgls.report' functionality of the *mmodely* package. ANOVA, AIC, and one-line model reports can be output via this function.

```
> model <- as.formula('OC ~ mass.Kg + group.size')
> fit <- caper::pgls(formula=model, data=comp)
> summary(fit)

Call:
caper::pgls(formula = model, data = comp)

Residuals:
    Min      1Q      Median      3Q      Max 
-55.382 -12.734  -4.075   27.512   57.101 

Branch length transformations:

kappa [Fix] : 1.000
lambda [Fix] : 1.000
delta [Fix] : 1.000

Coefficients:
            Estimate Std. Error t value Pr(>|t|)    
(Intercept) 66.037399   8.409402  7.8528 7.05e-07 ***
mass.Kg      0.153139   0.060894  2.5148  0.02298 *  
group.size   -0.030092   0.107433 -0.2801  0.78299    
---
Signif. codes:  0 ‘***’ 0.001 ‘**’ 0.01 ‘*’ 0.05 ‘.’ 0.1 ‘ ’ 1

Residual standard error: 34.2 on 16 degrees of freedom
Multiple R-squared: 0.3513,          Adjusted R-squared: 0.2702 
F-statistic: 4.332 on 2 and 16 DF,  p-value: 0.03136

> pgls.report(comp, f=model, anova=TRUE, QC.plot=TRUE)

Call:
pgls(formula = f, data = cd, lambda = l, kappa = k, delta = d,
     bounds = bounds)
```

Residuals:

	Min	1Q	Median	3Q	Max
	-55.382	-12.734	-4.075	27.512	57.101

Branch length transformations:

kappa [Fix] : 1.000
lambda [Fix] : 1.000
delta [Fix] : 1.000

Coefficients:

	Estimate	Std. Error	t value	Pr(> t)
(Intercept)	66.037399	8.409402	7.8528	7.05e-07 ***
mass.Kg	0.153139	0.060894	2.5148	0.02298 *
group.size	-0.030092	0.107433	-0.2801	0.78299

Signif. codes: 0 ‘***’ 0.001 ‘**’ 0.01 ‘*’ 0.05 ‘.’ 0.1 ‘ ’ 1

Residual standard error: 34.2 on 16 degrees of freedom

Multiple R-squared: 0.3513, Adjusted R-squared: 0.2702

F-statistic: 4.332 on 2 and 16 DF, p-value: 0.03136

[1] "AIC = 134.2"

Analysis of Variance Table

Sequential SS for pgls: lambda = 1.00, delta = 1.00, kappa = 1.00

Response: DC

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
mass.Kg	1	10043.9	10043.9	8.5862	0.009806 **
group.size	1	91.8	91.8	0.0785	0.782991
Residuals	16	18716.2	1169.8		

Signif. codes: 0 ‘***’ 0.001 ‘**’ 0.01 ‘*’ 0.05 ‘.’ 0.1 ‘ ’ 1

++mass(0.023) | group(0.783)

Call:

pgls(formula = f, data = cd, lambda = 1, kappa = k, delta = d,
bounds = bounds)

Coefficients:

(Intercept)	mass.Kg	group.size
66.03740	0.15314	-0.03009

6 Multivariate Combinatoric Iteration

The *mmodely* package's chief contribution is enabling approaches that utilize multi-model iteration averaging. Using a smaller subset of variables can speed up the (slower) maximum likelihood computation step and still achieve the desired result of fixed tree transformation parameters.

```
> pv0 <- c("mass.Kg", "group.size", "arboreal", "nocturnal") #'swing.pct"
> est.mods <- get.model.combos(predictor.vars=pv0, outcome.var='OC', min.q=2)
> ps <- get.phylo.stats(phylo=phyl, data=data, trait.clmn='OC');

$lambda
[1] 0.8133958

$logL
[1] -204.9656

$P
[1] 0

$K
[1] 0.7251836

$P
[1] 0.001

> lambda <- ps$lambda$lambda ; print(lambda)
[1] 0.8133958

> PGLSi <- pgls.iter(models=est.mods, phylo=phyl, df=data, l=lambda, k='ML', d='ML')

1 OC~mass.Kg+group.size+arboreal+nocturnal
2 OC~mass.Kg+group.size+arboreal
3 OC~mass.Kg+group.size+nocturnal
4 OC~mass.Kg+arboreal+nocturnal
5 OC~group.size+arboreal+nocturnal
6 OC~mass.Kg+group.size
7 OC~mass.Kg+arboreal
8 OC~mass.Kg+nocturnal
9 OC~group.size+arboreal
10 OC~group.size+nocturnal
11 OC~arboreal+nocturnal

> pgls.iter.stats(PGLSi) # check run, especially to see how few sub-datasets exist

models: 11
dimensions of sub-datasets:
    q      n   qXn rwGsm
    3      1     3     1
          q  n
Min. 2.000000 63
1st Qu. 2.000000 63
Median 2.000000 63
Mean   2.545455 63
3rd Qu. 3.000000 63
Max.   4.000000 63
tree transformation parameter averages:
    1           k           d
0.8133958 1.4591957 1.2008048
distributions of optimization parameters:
      n           q           rwGsm        model.no          R2
Min. :63   Min. :2.000   Min. :117909   Min. : 1.0   Min. :0.04241
```

1st Qu.:63	1st Qu.:2.000	1st Qu.:117909	1st Qu.: 3.5	1st Qu.:0.05945
Median :63	Median :2.000	Median :117909	Median : 6.0	Median :0.20084
Mean :63	Mean :2.545	Mean :117909	Mean : 6.0	Mean :0.15453
3rd Qu.:63	3rd Qu.:3.000	3rd Qu.:117909	3rd Qu.: 8.5	3rd Qu.:0.21835
Max. :63	Max. :4.000	Max. :117909	Max. :11.0	Max. :0.23433
R2.adj	AIC	AICc	BIC	AICw
Min. :0.01049	Min. :400.1	Min. :400.5	Min. :406.5	Min. :0.0003387
1st Qu.:0.02002	1st Qu.:401.5	1st Qu.:402.1	1st Qu.:409.3	1st Qu.:0.0006806
Median :0.16042	Median :402.6	Median :403.7	Median :412.0	Median :0.0654038
Mean :0.11865	Mean :405.8	Mean :406.4	Mean :413.4	Mean :0.0909091
3rd Qu.:0.18158	3rd Qu.:412.4	3rd Qu.:412.8	3rd Qu.:418.8	3rd Qu.:0.1460563
Max. :0.19116	Max. :413.5	Max. :414.2	Max. :422.1	Max. :0.3197358
BICw				
Min. :0.0001827				
1st Qu.:0.0009305				
Median :0.0287186				
Mean :0.0909091				
3rd Qu.:0.1117665				
Max. :0.4371383				

7 Tree Transformation Averaging and Re-iteration

After running PGLS on a test-subset of predictor-variable combinations using maximum likelihood, we can average the tree transformation parameters [18] to obtain fixed values going forward. This approach can speed up computations for larger sets of modeling data and variable combinations.

```
> tt.avgs <- apply(PGLSi$params, 2, mean, na.rm=TRUE) # tree transformation averages
> print(tt.avgs)
```

	1	k	d
0.8133958	1.4591957	1.2008048	

Next we use the full set of variables and our tree transform averages. For demonstration, we sprinkle in some missing values to our dataset so as to artificially boost the number of sub-datasets. The subsequent fixed tree parameter iteration run should now generate more diverse output upon which the *mmodely* can demonstrate its unique model averaging and model selection functionality.

```
> pvs <- c("mass.Kg", "group.size", "infant.carry", "arboreal", "DPL.km", "nocturnal")
> all.mods <- get.model.combos(predictor.vars=pvs, outcome.var='OC', min.q=2)
> data <- subset(data,!grepl(rownames(data),pattern='gorilla')) # remove an OC measurement outlier
> # randomly sprinkle in some missing values (for more interesting for model selection)
> missing.value.ct <- 1
> for(pv in pv0){ data[sample(x=1:nrow(data),size=missing.value.ct),pv] <- NA}
> PGLSi <- pgls.iter(models=all.mods, phylo=phyl, df=data, l=lambda, k=tt.avgs['k'], d=tt.avgs['d'])

1 OC~mass.Kg+group.size+infant.carry+arboreal+DPL.km+nocturnal
2 OC~mass.Kg+group.size+infant.carry+arboreal+DPL.km
3 OC~mass.Kg+group.size+infant.carry+arboreal+nocturnal
4 OC~mass.Kg+group.size+infant.carry+DPL.km+nocturnal
5 OC~mass.Kg+group.size+arboreal+DPL.km+nocturnal
6 OC~mass.Kg+infant.carry+arboreal+DPL.km+nocturnal
7 OC~group.size+infant.carry+arboreal+DPL.km+nocturnal
8 OC~mass.Kg+group.size+infant.carry+arboreal
9 OC~mass.Kg+group.size+infant.carry+DPL.km
10 OC~mass.Kg+group.size+infant.carry+nocturnal
11 OC~mass.Kg+group.size+arboreal+DPL.km
12 OC~mass.Kg+group.size+arboreal+nocturnal
13 OC~mass.Kg+group.size+DPL.km+nocturnal
14 OC~mass.Kg+infant.carry+arboreal+DPL.km
15 OC~mass.Kg+infant.carry+arboreal+nocturnal
16 OC~mass.Kg+infant.carry+DPL.km+nocturnal
17 OC~mass.Kg+arboreal+DPL.km+nocturnal
18 OC~group.size+infant.carry+arboreal+DPL.km
19 OC~group.size+infant.carry+arboreal+nocturnal
20 OC~group.size+infant.carry+DPL.km+nocturnal
21 OC~group.size+arboreal+DPL.km+nocturnal
22 OC~infant.carry+arboreal+DPL.km+nocturnal
23 OC~mass.Kg+group.size+infant.carry
24 OC~mass.Kg+group.size+arboreal
25 OC~mass.Kg+group.size+DPL.km
26 OC~mass.Kg+group.size+nocturnal
27 OC~mass.Kg+infant.carry+arboreal
28 OC~mass.Kg+infant.carry+DPL.km
29 OC~mass.Kg+infant.carry+nocturnal
30 OC~mass.Kg+arboreal+DPL.km
31 OC~mass.Kg+arboreal+nocturnal
32 OC~mass.Kg+DPL.km+nocturnal
33 OC~group.size+infant.carry+arboreal
34 OC~group.size+infant.carry+DPL.km
35 OC~group.size+infant.carry+nocturnal
36 OC~group.size+arboreal+DPL.km
37 OC~group.size+arboreal+nocturnal
```

```

38 OC~group.size+DPL.km+nocturnal
39 OC~infant.carry+arboreal+DPL.km
40 OC~infant.carry+arboreal+nocturnal
41 OC~infant.carry+DPL.km+nocturnal
42 OC~arboreal+DPL.km+nocturnal
43 OC~mass.Kg+group.size
44 OC~mass.Kg+infant.carry
45 OC~mass.Kg+arboreal
46 OC~mass.Kg+DPL.km
47 OC~mass.Kg+nocturnal
48 OC~group.size+infant.carry
49 OC~group.size+arboreal
50 OC~group.size+DPL.km
51 OC~group.size+nocturnal
52 OC~infant.carry+arboreal
53 OC~infant.carry+DPL.km
54 OC~infant.carry+nocturnal
55 OC~arboreal+DPL.km
56 OC~arboreal+nocturnal
57 OC~DPL.km+nocturnal

```

8 Fixed iteration run statistics

We should briefly inspect how this fixed iteration run performed and how many sub-datasets we need to investigate. It is recommended to try *mmodely* using 'rwGsm.' This abbreviation stands for 'raw *Genus species* sums.' It represents a sum of the (concatenated) raw character values of all species constituting the underlying dataset (which has all rows with any missing data removed) for a particular combination of model predictor variables. While this default is preferred, the number of species 'n' [default] or number of model variables 'q' can also be used.

```

> pgls.iter.stats(PGLSi)

models: 57
dimensions of sub-datasets:
      q      n    qXn rwGsm
      5      9     16     31
      q      n
Min.   2.000000 53.00000
1st Qu. 2.000000 55.00000
Median  3.000000 56.00000
Mean    3.263158 57.10526
3rd Qu. 4.000000 60.00000
Max.   6.000000 61.00000
tree transformation parameter averages:
      1      k      d
0.8133958 1.4591957 1.2008048
distributions of optimization parameters:
      n      q      rwGsm      model.no      R2
Min. :53.00  Min. :2.000  Min. : 98558  Min. : 1  Min. :0.00362
1st Qu.:55.00 1st Qu.:2.000  1st Qu.:102113 1st Qu.:15  1st Qu.:0.06607
Median :56.00  Median :3.000  Median :104197  Median :29  Median :0.10661
Mean   :57.11  Mean   :3.263  Mean   :106654  Mean   :29  Mean   :0.09621
3rd Qu.:60.00  3rd Qu.:4.000  3rd Qu.:112386  3rd Qu.:43  3rd Qu.:0.11997
Max.   :61.00  Max.   :6.000  Max.   :114596  Max.   :57  Max.   :0.16022
      R2.adj      AIC      AICc      BIC      AICw
Min. :-0.03074  Min. :345.9  Min. :347.8  Min. :357.8  Min. :0.0000000
1st Qu.: 0.02009 1st Qu.:356.4  1st Qu.:357.4  1st Qu.:365.0  1st Qu.:0.0000000
Median : 0.04045  Median :364.5  Median :365.3  Median :372.6  Median :0.0000738
Mean   : 0.03995  Mean   :369.9  Mean   :370.8  Mean   :378.5  Mean   :0.0175439
3rd Qu.: 0.06142  3rd Qu.:384.6  3rd Qu.:385.7  3rd Qu.:392.6  3rd Qu.:0.0038459
Max.   : 0.09923  Max.   :395.4  Max.   :395.8  Max.   :401.7  Max.   :0.4671129

```

```
BICw
Min. :0.0000000
1st Qu.:0.0000000
Median :0.0002093
Mean   :0.0175439
3rd Qu.:0.0092162
Max.   :0.3472579
```

9 Model Averaging

Now we can estimate the predictor variable parameters by averaging over all possible fixed PGLS runs, using the AICc differences (from the lowest AICc) as weights. By default this AICw weighted average is performed per sub-dataset using 'rwGsm' or 'n' [default] as mentioned in the preceding section. While model averaging is not recommended under high multicollinearity, as denominators of regression coefficients change across models, it is possible to rescale these using 'standardize' [13]. A slightly more conservative alternative to MA uses 'model importance' which is equivalent to an AIC-weighted MA of binary indicators of presence or absence of covariate model inclusion [12].

```
> w.means.pds <- average.fit.models(vars=pvs, fits=PGLSi$fits, optims=PGLSi$optim, by='rwGsm', standardize=TRUE)
> #
> apply(w.means.pds, 2, mean, na.rm=T) #average of weighted means over all sub-datasets

      mass.Kg group.size infant.carry      arboreal      DPL.km    nocturnal
0.14808937  0.09737625  5.42210437  0.58122187 -0.14692097 -1.97411938

> w.means.pds
# weighted means      per sub-dataset

      mass.Kg group.size infant.carry      arboreal      DPL.km    nocturnal
98558  0.15703  0.09432  5.10026  2.15505 -0.10861 -1.44005
100270 0.17071        NaN  5.56540  1.20656  0.55895 -1.62289
100275 0.13427  0.08808  4.60875        NaN -0.57430 -1.24644
100396 0.15618  0.09391  5.16899  2.15042 -0.01737        NaN
100642        NaN  0.09954  5.46251  1.34996 -0.23355 -2.61006
101987 0.15675        NaN  5.19131        NaN  0.15665 -1.52027
102108 0.17006        NaN  5.61600  1.18237  0.63972        NaN
102113 0.13306  0.08664  4.75636        NaN -0.51235        NaN
102354        NaN        NaN  5.93049  0.22829  0.51250 -2.68039
102359        NaN  0.09643  5.09970        NaN -0.52894 -2.48492
102480        NaN  0.09957  5.60673  1.25806 -0.08883        NaN
103825 0.15623        NaN  5.27714        NaN  0.21657        NaN
104071        NaN        NaN  5.75197        NaN  0.31787 -2.66272
104192        NaN        NaN  6.07194  0.12960  0.63980        NaN
104197        NaN  0.09671  5.29904        NaN -0.40059        NaN
105909        NaN        NaN  6.24708        NaN  0.44010        NaN
108957 0.13540  0.09444        NaN  0.66988 -0.66177 -1.16128
110669 0.15206        NaN        NaN -0.18125 -0.09096 -1.36769
110674 0.12731  0.09724        NaN        NaN -0.72107 -1.08671
110795 0.13532  0.09349        NaN  0.79417 -0.61195        NaN
111041        NaN  0.09968        NaN  0.11684 -0.71865 -2.51637
112386 0.15303        NaN        NaN        NaN -0.06473 -1.40119
112507 0.15207        NaN        NaN -0.08382 -0.04130        NaN
112512 0.12620  0.09537        NaN        NaN -0.69933        NaN
112753        NaN        NaN        NaN -0.90045 -0.08380 -2.61947
112758        NaN  0.10529        NaN        NaN -0.68907 -2.47073
112879        NaN  0.09962        NaN  0.13621 -0.62789        NaN
114224 0.15375        NaN        NaN        NaN -0.03589        NaN
114470        NaN        NaN        NaN        NaN  0.08042 -2.69473
114591        NaN        NaN        NaN -0.91234  0.00561        NaN
114596        NaN  0.11769        NaN        NaN -0.61179        NaN
attr(", "MSE")
      mass.Kg group.size infant.carry      arboreal      DPL.km    nocturnal
```

98558	0.00057	0.00163	0.03826	0.04211	0	0.01898
100270	0.00006	NA	0.02798	0.02942	0	0.00602
100275	0.00039	0.00151	0.05094	NA	0	0.01721
100396	0.00021	0.00018	0.00430	0.00059	0	NA
100642	NA	0.00012	0.00462	0.00434	0	0.00152
101987	0.00001	NA	0.00136	NA	0	0.00039
102108	0.00001	NA	0.00792	0.00861	0	NA
102113	0.00010	0.00035	0.01155	NA	0	NA
102354	NA	NA	0.00088	0.00090	0	0.00014
102359	NA	0.00007	0.00228	NA	0	0.00075
102480	NA	0.00001	0.00017	0.00026	0	NA
103825	0.00000	NA	0.00042	NA	0	NA
104071	NA	NA	0.00008	NA	0	0.00002
104192	NA	NA	0.00017	0.00018	0	NA
104197	NA	0.00001	0.00027	NA	0	NA
105909	NA	NA	0.00000	NA	0	NA
108957	0.00000	0.00000	NA	0.00000	0	0.00000
110669	0.00000	NA	NA	0.00000	0	0.00000
110674	0.00000	0.00000	NA	NA	0	0.00000
110795	0.00000	0.00000	NA	0.00000	0	NA
111041	NA	0.00000	NA	0.00000	0	0.00000
112386	0.00000	NA	NA	NA	0	0.00000
112507	0.00000	NA	NA	0.00000	0	NA
112512	0.00000	0.00000	NA	NA	0	NA
112753	NA	NA	NA	0.00000	0	0.00000
112758	NA	0.00000	NA	NA	0	0.00000
112879	NA	0.00000	NA	0.00000	0	NA
114224	0.00000	NA	NA	NA	0	NA
114470	NA	NA	NA	NA	0	0.00000
114591	NA	NA	NA	0.00000	0	NA
114596	NA	0.00000	NA	NA	0	NA

```
> w.import.pds <- variable.importance(vars=pvs, fits=PGLSi$fits, optims=PGLSi$optim, by='rwGsm')
> #
> apply(w.import.pds, 2, mean, na.rm=T) #average of weighted means over all sub-datasets
```

mass.Kg	group.size	infant.carry	arboreal	DPL.km	nocturnal
1.0000000	1.0000000	1.0000000	0.6693031	0.3225806	0.0000000

```
> w.import.pds # weighted means per sub-dataset
```

mass.Kg	group.size	infant.carry	arboreal	DPL.km	nocturnal
98558	1	1	1	1.00000	0
100270	1	NaN	1	1.00000	1
100275	1	1	1	NaN	0
100396	1	1	1	1.00000	0
100642	NaN	1	1	1.00000	0
101987	1	NaN	1	NaN	1
102108	1	NaN	1	1.00000	1
102113	1	1	1	NaN	0
102354	NaN	NaN	1	1.00000	1
102359	NaN	1	1	NaN	0
102480	NaN	1	1	1.00000	0
103825	1	NaN	1	NaN	1
104071	NaN	NaN	1	NaN	1
104192	NaN	NaN	1	0.27372	1
104197	NaN	1	1	NaN	0
105909	NaN	NaN	1	NaN	1
108957	1	1	NaN	1.00000	0
110669	1	NaN	NaN	0.00000	0
110674	1	1	NaN	NaN	0

110795	1	1	NaN	1.00000	0	NaN
111041	NaN	1	NaN	0.71607	0	0
112386	1	NaN	NaN	NaN	0	0
112507	1	NaN	NaN	0.00000	0	NaN
112512	1	1	NaN	NaN	0	NaN
112753	NaN	NaN	NaN	0.00000	0	0
112758	NaN	1	NaN	NaN	0	0
112879	NaN	1	NaN	0.71906	0	NaN
114224	1	NaN	NaN	NaN	0	NaN
114470	NaN	NaN	NaN	NaN	1	0
114591	NaN	NaN	NaN	0.00000	1	NaN
114596	NaN	1	NaN	NaN	0	NaN
attr(,"MSE")						
	mass.Kg	group.size	infant.carry	arboreal	DPL.km	nocturnal
98558	0	0	0	0.00000	0	0
100270	0	NA	0	0.00000	0	0
100275	0	0	0	NA	0	0
100396	0	0	0	0.00000	0	NA
100642	NA	0	0	0.00000	0	0
101987	0	NA	0	NA	0	0
102108	0	NA	0	0.00000	0	NA
102113	0	0	0	NA	0	NA
102354	NA	NA	0	0.00000	0	0
102359	NA	0	0	NA	0	0
102480	NA	0	0	0.00000	0	NA
103825	0	NA	0	NA	0	NA
104071	NA	NA	0	NA	0	0
104192	NA	NA	0	0.00022	0	NA
104197	NA	0	0	NA	0	NA
105909	NA	NA	0	NA	0	NA
108957	0	0	NA	0.00000	0	0
110669	0	NA	NA	0.00000	0	0
110674	0	0	NA	NA	0	0
110795	0	0	NA	0.00000	0	NA
111041	NA	0	NA	0.00000	0	0
112386	0	NA	NA	NA	0	0
112507	0	NA	NA	0.00000	0	NA
112512	0	0	NA	NA	0	NA
112753	NA	NA	NA	0.00000	0	0
112758	NA	0	NA	NA	0	0
112879	NA	0	NA	0.00000	0	NA
114224	0	NA	NA	NA	0	NA
114470	NA	NA	NA	NA	0	0
114591	NA	NA	NA	0.00000	0	NA
114596	NA	0	NA	NA	0	NA

10 Model Selection

We can select the best model by sorting each subset (e.g. by AICc) or by using visualization methods.

```
> select.best.models(PGLSi, using='AICc')
```

n	q	qXn	rwGsm	model.no	R2	R2.adj	AIC	AICc	BIC	AICw
53	53	5	5X53	98558	3	0.16007974	0.07072652	345.9441	347.7702	357.7659
54	54	4	4X54	100396	8	0.15679638	0.08796344	349.0633	350.3133	359.0083
55	55	3	3X55	102113	23	0.14927592	0.09923333	353.2706	354.0706	361.2999
56	56	2	2X56	103825	44	0.11006887	0.07648657	358.7657	359.2273	364.8418
57	57	2	2X57	105909	53	0.05921713	0.02437332	368.4776	368.9304	374.6067
58	58	4	4X58	108957	12	0.10716908	0.03978561	376.1636	377.3174	386.4658
59	59	3	3X59	110795	24	0.10155643	0.05255042	379.4725	380.2132	387.7826
60	60	2	2X60	112512	43	0.10755127	0.07623728	382.9089	383.3374	389.1919
61	61	2	2X61	114224	46	0.05993341	0.02751732	391.0582	391.4792	397.3908
										1.507064e-10
										BICw
53										3.472579e-01
54										1.865819e-01
55										5.932599e-02
56										1.009574e-02
57										7.650832e-05
58										2.034870e-07
59										1.053416e-07
60										5.206880e-08
61										8.633999e-10

Plotting the coefficients of determination versus the AIC values allows selection of high-performing models for inspection and reporting.

```
> plot.pgls.iters(PGLSi)
```

```
> sdevs.objs <- get.pgls.coefs(PGLSi$fits, est='t value')
> coefs.objs <- get.pgls.coefs(PGLSi$fits, est='Estimate')

> report.vect <- sapply(1:length(PGLSi$fits), function(i) fit.1ln.rprt(PGLSi$fits[[i]], rtrn.line=FALSE, mn=i))

1 +mass(0.108) +group(0.162) +infan(0.204) arbore(0.452) | DPL(0.93) noctur(0.598) R2adj: 0.051 AICc: 350.42
2 +mass(0.097) +group(0.156) +infan(0.168) arbore(0.412) | DPL(0.988) R2adj: 0.069 AICc: 352.85
3 +mass(0.102) +group(0.133) +infan(0.153) arbore(0.369) | noctur(0.598) R2adj: 0.071 AICc: 347.77
4 +group(0.125) +mass(0.134) +infan(0.261) | noctur(0.604) DPL(0.583) R2adj: 0.071 AICc: 353.53
5 +group(0.095) +mass(0.133) arbore(0.917) | noctur(0.636) DPL(0.514) R2adj: 0.029 AICc: 379.33
6 +mass(0.072) +infan(0.11) arbore(0.516) DPL(0.612) | noctur(0.581) R2adj: 0.032 AICc: 355.07
7 +group(0.131) +infan(0.181) arbore(0.66) | DPL(0.851) -noctu(0.31) R2adj: 0.031 AICc: 356.65
8 +mass(0.093) +group(0.12) +infan(0.131) arbore(0.348) | R2adj: 0.088 AICc: 350.31
9 +group(0.123) +mass(0.125) +infan(0.219) | DPL(0.614) R2adj: 0.086 AICc: 356.21
10 +mass(0.14) +group(0.144) +infan(0.17) | noctur(0.649) R2adj: 0.084 AICc: 351.34
11 +group(0.091) +mass(0.121) arbore(0.861) | DPL(0.536) R2adj: 0.042 AICc: 382.18
12 +group(0.12) +mass(0.131) arbore(0.714) | noctur(0.677) R2adj: 0.04 AICc: 377.32
13 +group(0.064) +mass(0.124) | noctur(0.631) DPL(0.427) R2adj: 0.06 AICc: 381.97
14 +mass(0.064) +infan(0.089) arbore(0.485) DPL(0.549) | R2adj: 0.05 AICc: 357.63
15 +mass(0.069) +infan(0.126) arbore(0.649) | noctur(0.534) R2adj: 0.046 AICc: 352.83
16 +mass(0.082) +infan(0.135) DPL(0.863) | noctur(0.579) R2adj: 0.044 AICc: 358.72
17 +mass(0.086) | DPL(0.923) arbore(0.914) noctur(0.605) R2adj: -0.004 AICc: 385.02
18 +group(0.13) +infan(0.142) arbore(0.64) | DPL(0.942) R2adj: 0.033 AICc: 360.08
19 +group(0.113) +infan(0.123) arbore(0.551) | -noctu(0.312) R2adj: 0.05 AICc: 354.16
20 +group(0.095) +infan(0.203) | DPL(0.616) -noctu(0.302) R2adj: 0.058 AICc: 359.51
21 +group(0.077) | arbore(0.891) DPL(0.484) -noctu(0.305) R2adj: 0.011 AICc: 385.75
22 +infan(0.093) DPL(0.647) arbore(0.784) | -noctu(0.315) R2adj: 0.005 AICc: 361.81
23 +mass(0.13) +group(0.134) +infan(0.141) | R2adj: 0.099 AICc: 354.07
24 +group(0.113) +mass(0.121) arbore(0.662) | R2adj: 0.053 AICc: 380.21
```

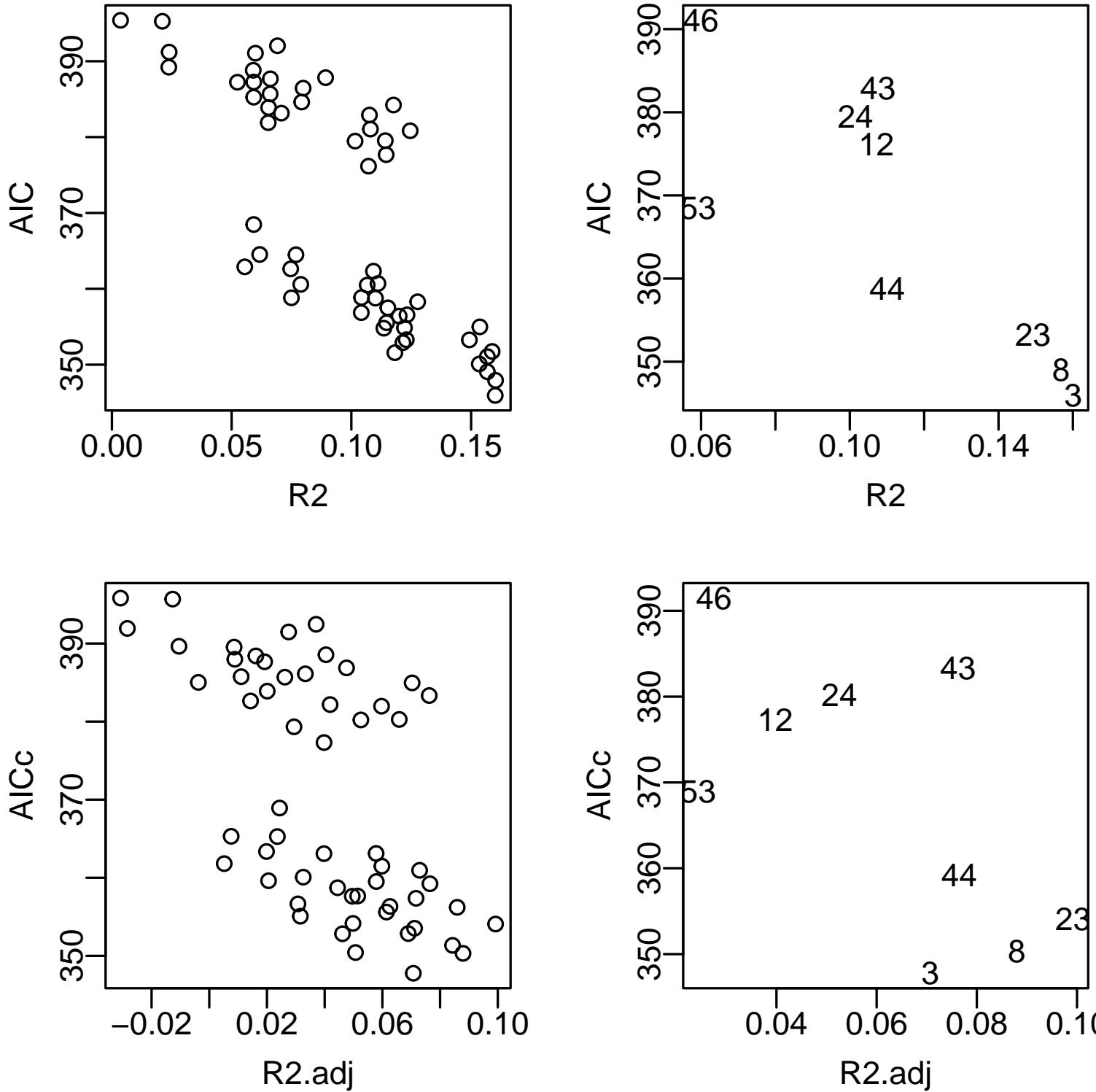


Figure 1: All possible model combinations appear as individual points above. As there is a generally negative association between AIC and the coefficient of determination, the points tend to follow a negative sloping streak to the lower right. The "best" models typically appear in the lower right of each streak. Therefore, minimizing AIC tends to also maximize the coefficient of determination, but not necessarily. This four panel plot looks at correct and adjusted versions of each model assessment measure. All points are scaled by subdataset sample size by default if 'n' is used in grouping.

```

25 +group(0.062) +mass(0.117) | DPL(0.428) R2adj: 0.07 AICc: 384.96
26 +group(0.091) +mass(0.138) | noctur(0.695) R2adj: 0.066 AICc: 380.27
27 +mass(0.062) +infan(0.108) arbore(0.651) | R2adj: 0.061 AICc: 355.61
28 +mass(0.075) +infan(0.111) DPL(0.805) | R2adj: 0.06 AICc: 361.48
29 +mass(0.071) +infan(0.13) | noctur(0.552) R2adj: 0.063 AICc: 356.33
30 +mass(0.077) | DPL(0.964) arbore(0.959) R2adj: 0.009 AICc: 387.99
31 +mass(0.083) | arbore(0.941) noctur(0.607) R2adj: 0.014 AICc: 382.64
32 +mass(0.068) | DPL(0.938) noctur(0.585) R2adj: 0.016 AICc: 388.41
33 +group(0.102) +infan(0.103) arbore(0.569) | R2adj: 0.051 AICc: 357.66
34 +group(0.095) +infan(0.16) | DPL(0.698) R2adj: 0.058 AICc: 363.11
35 +group(0.099) +infan(0.129) | noctur(0.324) R2adj: 0.072 AICc: 357.37
36 +group(0.075) | arbore(0.912) DPL(0.534) R2adj: 0.009 AICc: 389.54
37 +group(0.101) arbore(0.894) | noctur(0.328) R2adj: 0.02 AICc: 383.9
38 ++grou(0.046) | DPL(0.451) -noctu(0.292) R2adj: 0.04 AICc: 388.56
39 +infan(0.071) DPL(0.559) arbore(0.775) | R2adj: 0.008 AICc: 365.31
40 +infan(0.102) arbore(0.976) | -noctu(0.285) R2adj: 0.021 AICc: 359.62
41 +infan(0.092) DPL(0.726) | -noctu(0.303) R2adj: 0.024 AICc: 365.28
42 | DPL(0.93) arbore(0.678) -noctu(0.299) R2adj: -0.028 AICc: 391.93
43 +group(0.088) +mass(0.131) | R2adj: 0.076 AICc: 383.34
44 +mass(0.062) +infan(0.11) | R2adj: 0.076 AICc: 359.23
45 +mass(0.075) | arbore(0.971) R2adj: 0.026 AICc: 385.69
46 +mass(0.062) | DPL(0.964) R2adj: 0.028 AICc: 391.48
47 +mass(0.064) | noctur(0.586) R2adj: 0.033 AICc: 386.12
48 +group(0.086) +infan(0.105) | R2adj: 0.073 AICc: 360.95
49 +group(0.092) arbore(0.891) | R2adj: 0.019 AICc: 387.66
50 ++grou(0.044) | DPL(0.495) R2adj: 0.037 AICc: 392.46
51 +group(0.061) | noctur(0.324) R2adj: 0.048 AICc: 386.88
52 +infan(0.085) | arbore(0.964) R2adj: 0.02 AICc: 363.36
53 +infan(0.071) DPL(0.619) | R2adj: 0.024 AICc: 368.93
54 +infan(0.096) | -noctu(0.272) R2adj: 0.04 AICc: 363.09
55 DPL(0.995) | arbore(0.688) R2adj: -0.031 AICc: 395.82
56 | arbore(0.671) -noctu(0.297) R2adj: -0.01 AICc: 389.64
57 DPL(0.923) | -noctu(0.279) R2adj: -0.013 AICc: 395.69

> par(mar=c(5,5,3,3))
> plot.pgls.R2AIC(PGLSi$optim)

```

11 Coefficient Plotting

Finally, the resulting model fits from the PGLS runs can be plotted out horizontally as distributions so the influence of each ecological predictor variable can be compared.

```

> par.old <- par(mar=c(5,8,1,4),mfrow=c(2,1))
> sparge.modsel(sdevs.objs, R2x=7, xlab='t value')
> sparge.modsel(coefs.objs, R2x=7, xlab='Estimate')

```

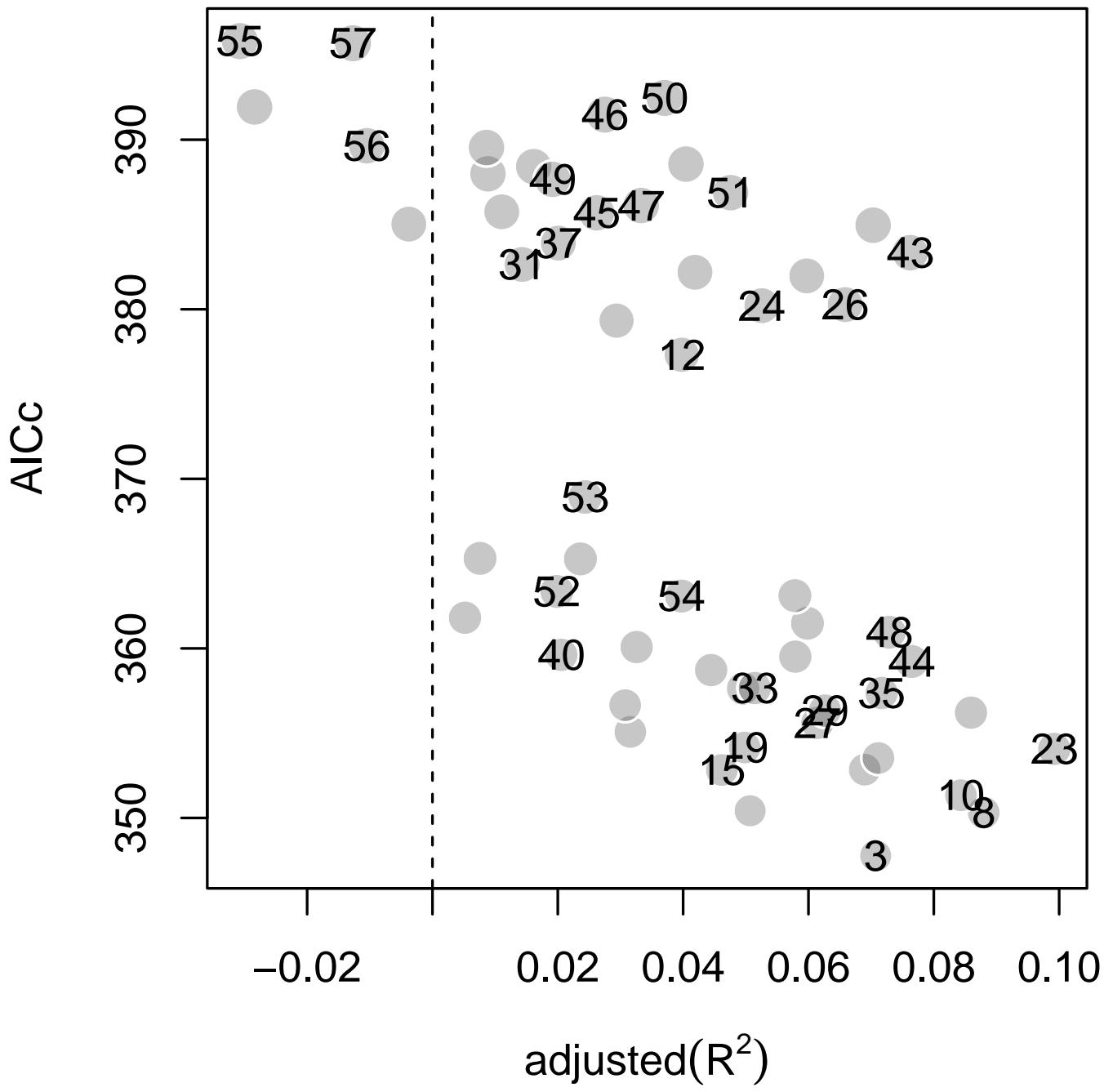


Figure 2: This is a one panel version of the previous model selection plot. The numbered points in the lower right corner of each streak of possible models represent the best model within a sub-dataset. Since these AICc values should not strictly be compared, it is recommended that all "best" models selected from each sub-dataset should be inspected or reported somehow, such as in the form of the sparge plot below.

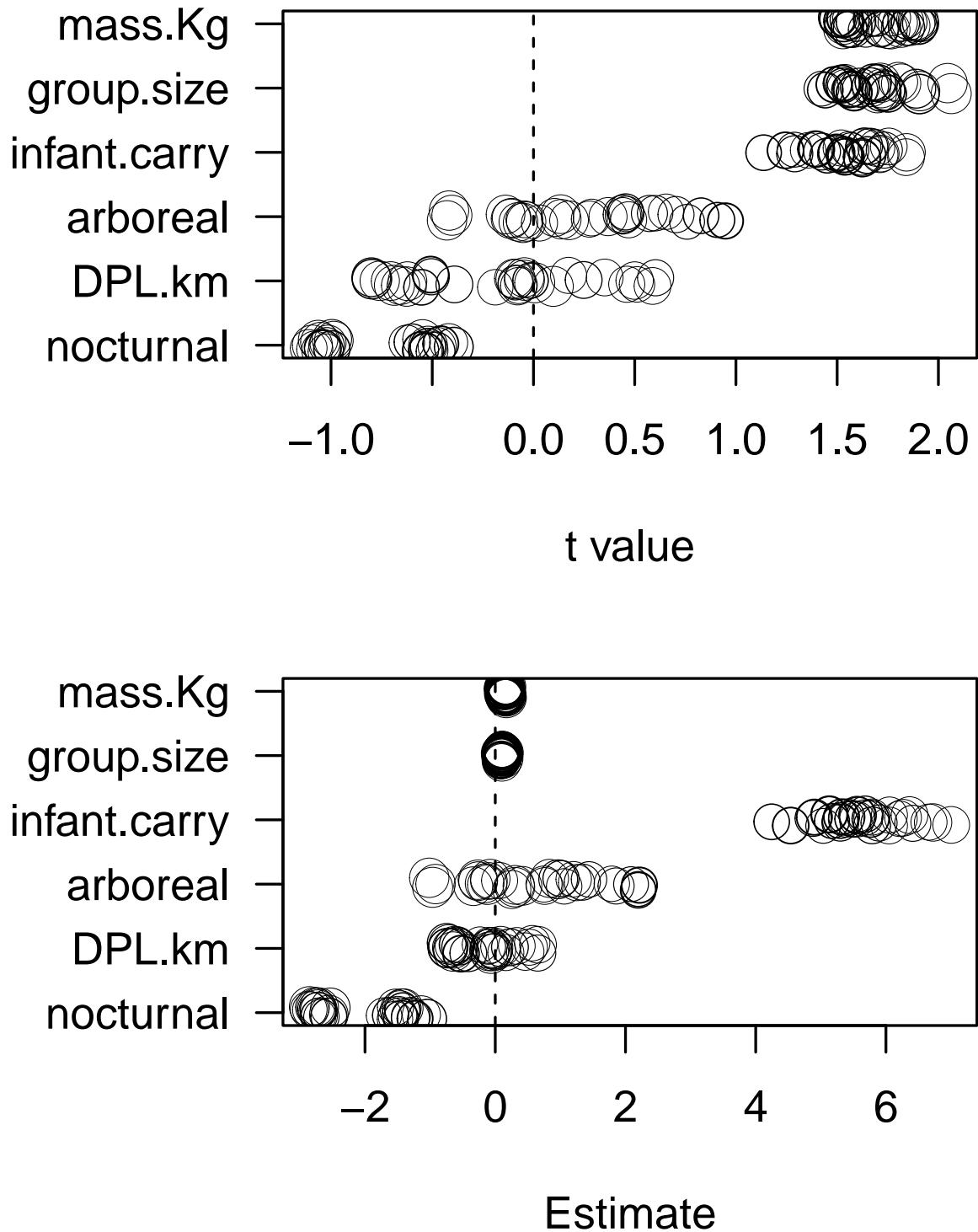


Figure 3: These horizontal parameter distribution dots plots demonstrate how the (t-values of) coefficients from all models can be simultaneously plot in order to verify consistency of estimates across the various (often missing-data driven) sub-datasets. To visually assess potential over-fitting of each model, point sizes represent underlying sample sizes and circle thickness corresponds to coefficient of determination values. Note that arboreal locomotor targeting and staturaly protective (e.g. infant fur-cling carrying) factors play key roles driving orbital convergence.

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